Light Brown Apple Moth in Nurseries

Steve Tjosvold University of California Cooperative Extension

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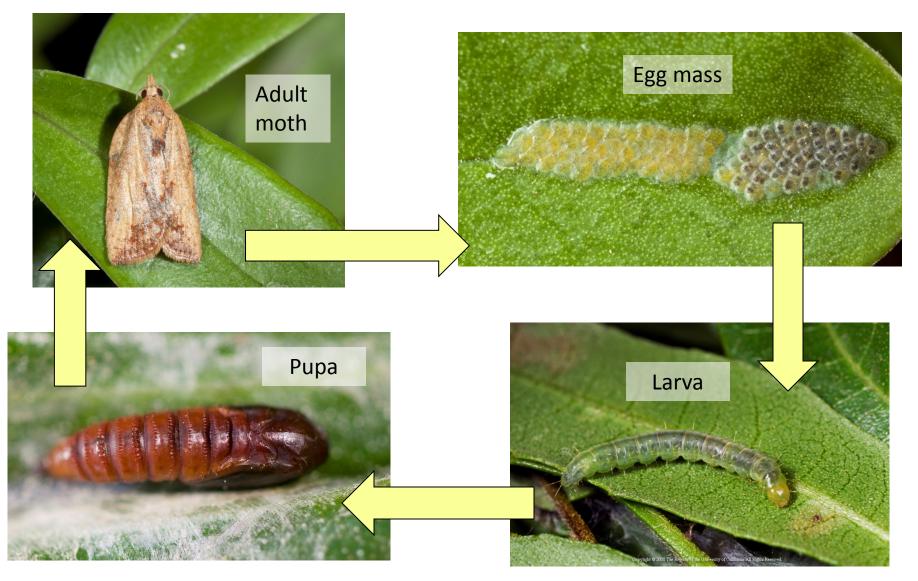
Light Brown Apple Moth (LBAM)

- Invasive pest from Australia detected in Alameda Co. in March 2007
- Favored by cooler coastal climates (20 counties)
- Broad host range (Established in natural areas and landscapes)
- Production loss due to regulatory closures
- Great increase in pesticide use in nurseries



December, 2013

LBAM life stages and cycle



No diapause, development continues 45 °F to 88 °F. Nearly 4 life cycles in Watsonville

Foliar Damage



Leaves chewed



Leaves distorted



Leaves bound together with silk-like webs





Fruit Damage

Family: Tortricidae Leafroller adults

- Wings bell-shaped
- Protruding mouthparts
- Antennae threadlike
- 0.25-1.25 in. wingspan
- Gray, tan, or brown with dark bands, mottled areas, metallic spots



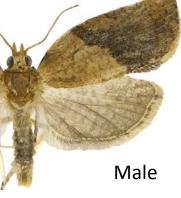




LBAM

Males : $\approx 8 \text{ mm}$ (0.3 in.) Female: $\approx 10 \text{ mm}$ (0.4 in.)

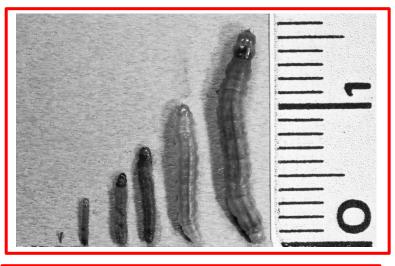






LBAM Larvae Field Identification

- 5 or 6 larva instars. Range in size from
 2 mm to 18 mm (up to 0.75 inch)
- Light to medium green body
- Light yellow-brown head
- White hairs and light legs.
- Body darker on top.
- 3 distinct darker bands running the length of the body
- Prothorax shield light green-brown
- Pre-spiracular pinaculum (3 hairs) and anal comb (7 or 8 prongs)









Not LBAM





DNA testing of suspected LBAM larvae is often necessary for identification in official detections

HOST PLANTS

545 plant species, 363 genera, and 121 families* Herbaceous plants preferred over woody plants Common: many weeds, ornamentals, and berry crops

Adiantum sp., Aquilegia sp., Amaranthus sp., Arbutus sp., apple (Malus domestica, Malus spp.), apricot (Prunus armeniaca), Artemesia sp., Astartea sp., Aster sp., avocado (Persea americana), Baccharis sp., black alder/European alder (Alnus glutinosa), blackberry and raspberry (Rubus spp.), black poplar (Populus nigra), blueberry (Vaccinium sp.), Boronia sp., Brassica sp., Breynia sp., broad bean (Vicia faba), broadleaf dock (Rumex obtusifolius), Bursaria sp., butterfly bush (Buddleia sp.), Calendula sp., Callistemon sp., camellia (Camellia japonica), Campsis sp., capeweed (Arctotheca calendula), Cassia sp., Ceanothus sp., Chinese gooseberry (Actinidia chinensis), Choisya sp., chrysanthemum (Chrysanthemum sp.), citrus (Citrus spp.), Clematis sp., Correa sp., cotoneaster (Cotoneaster sp.), Clerodendron sp., clover (Trifolium repens, Trifolium sp.), Cupressus sp., curled dock (Rumex crispus), currant (Ribes sp.), Cydonia sp., Dahlia sp., Datura sp., Daucus sp., Dodonaea sp., Eriobotrya sp., Eriostemon sp., Escallonia sp., eucalyptus (Eucalyptus sp.), euonymus (Euonymus sp.), fat-hen (Chenopodium album), Forsythia sp., Fortunella sp., fox's brush (Centranthus spp.), Gelsemium sp., Genista sp., Gerbera sp., gorse (Ulex europaeus), grape (Vitis vinifera, Vitis sp.), Grevillea sp., Hardenbergia sp., hawthorn (Crataegus sp.), hebe (Hebe spp.), Helichrysum sp., hop (Humulus lupulus), horn of plenty (Feijoa sellowiana), ivy (Hedera helix, Hedera spp.), jasmine (Jasminum spp.), Juglans sp., kiwifruit (Actinidia deliciosa), Lathyrus sp., Lavendula sp., Leucodendron sp., Leptospermum sp., Linus sp., litchi (Litchi chinensis), Lonicera sp., alfalfa (Medicago sativa), Lupinus sp., Lycopersicum sp., Macadamia sp., malabar ebony (Diospyros sp.), Mangifera sp., Melaleuca sp., Mentha sp., Mesembryanthemum sp., Michelia sp., Monotoca sp., montbretia (Crocosmia sp.), Myoporum sp., oak (Quercus sp.), Oxalis sp., Parthenocissus sp., peach (Prunus persica), pear (Pyrus sp.), Pelargonium sp., Persoonia sp., Petroselinum sp., persimmon (Diospyros kaki), Philadelphus sp., Photinia sp., Pittosporum sp., pine (Pinus muricata, P. radiata, Pinus sp.), plantain / ribwort (Plantago lanceolata), Platysace sp., Polygala sp., Polygonum sp., poplar and cottonwood (Populus nigra, Populus sp.), potato (Solanum tuberosum), privet (Ligustrum vulgare, Ligustrum sp.), Pteris sp., Pulcaria sp., Pyllanthus sp, Pyracantha sp., Ranunculus sp., Raphanus sp., Reseda sp., raspberry and boysenberry (Rubus idaeus, Rubus sp.), rose (Rosa sp.), Salvia sp., Senecio sp., Scotch broom (Cytisus scoparius), Sida sp., Sisymbrium sp., Smilax sp., Sollya sp., St. John's wort (Hypericum perforatum), strawberry (Fragaria sp.), Tithonia sp., Trema sp., Triglochin sp., Urtica sp., Viburnum sp., Vinca sp., wattle (Acacia sp.), willow (Salix sp.).

*E.G. Brokerhoff, et.al. (2011)

Eradication in Nurseries is Difficult LBAM can be introduced from surroundings

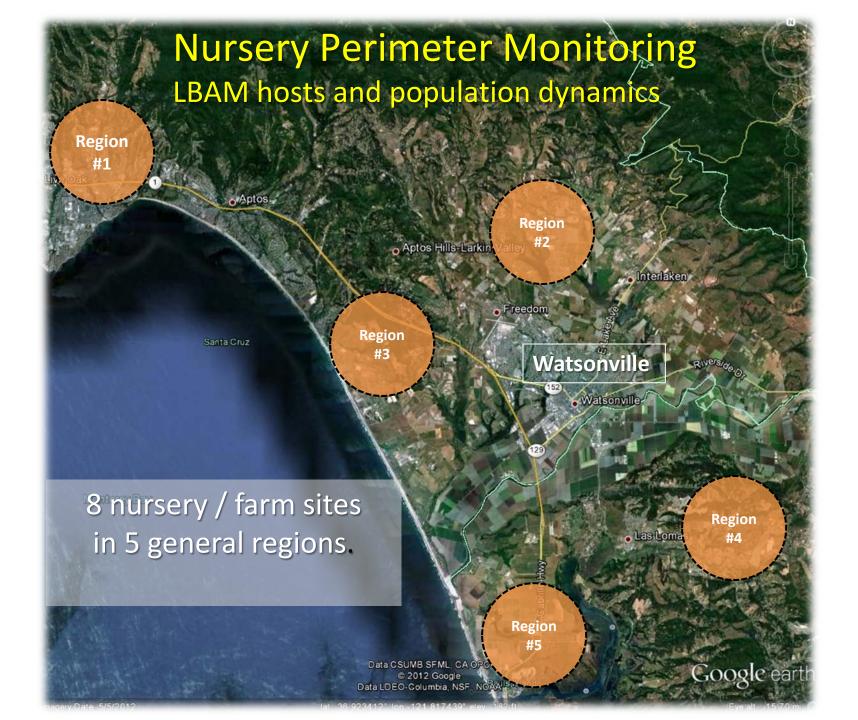


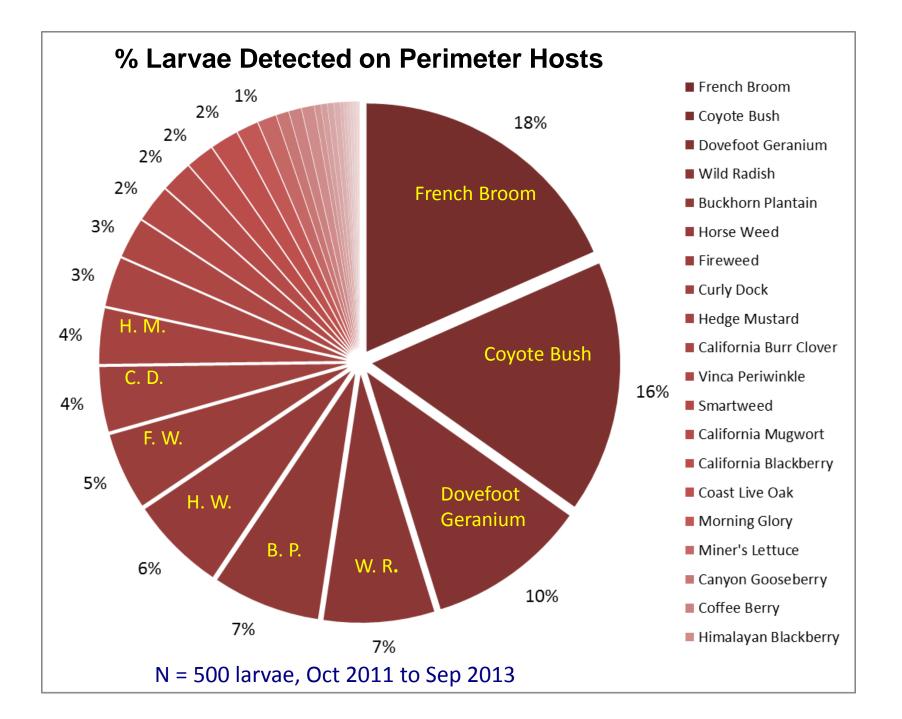


LBAM Larvae Detection in Raspberries (1A)

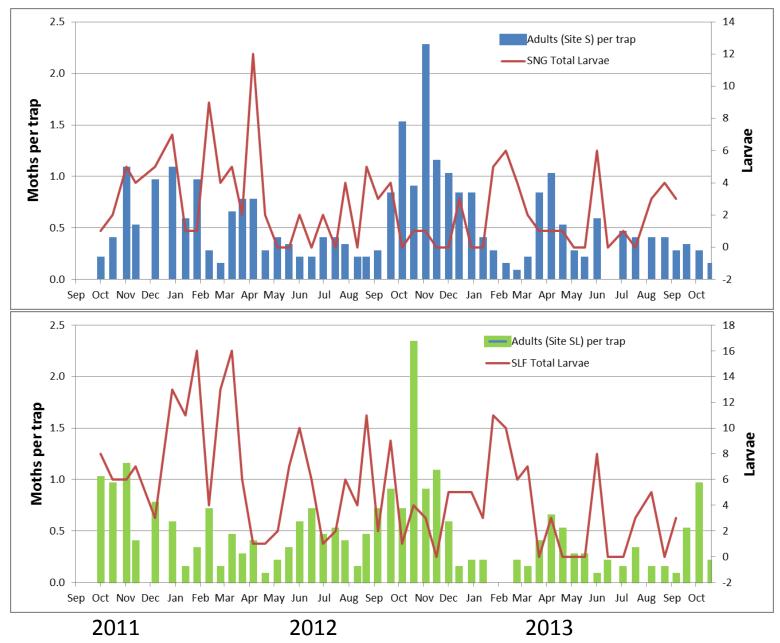


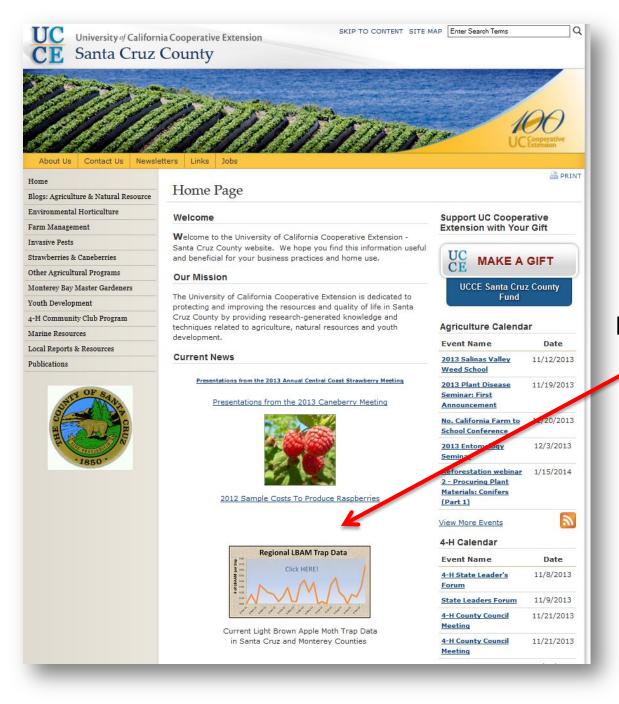
RED number of leaf rolls **BLUE** number of larvae





Moth and Larvae Numbers (2 of 8 Sites)





UCCE Santa Cruz home page

http:// cesantacruz.ucanr.edu

LBAM Parasitoids Indigenous in California

Trichogramma platneri and *T. fasciatum* Egg parasitoids



Meteorus trachynotus Larval parasitoid



Enytus eureka Larval parasitoid





Slide adapted from: Nick Mills, UC Berkeley

Biological Control

Augment biological control with releases of commercially available *Trichogramma platneri* at nursery perimeters or within nursery.

Bill Roltsch, CDFA







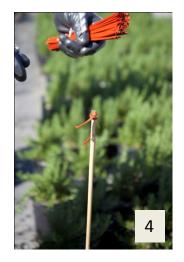
Isomate® LBAM Plus Application in Nurseries











300 twist ties per Acre = 13 X 13 foot square

Insecticide Control and Residual Activity

Treatment	% Control	Residual
Dipel (Bacillus thuringiensis) Provaunt (indoxacarb)	61 to 93 %	1 week
Conserve (spinosad)	82 to 100 %	1 to 2 weeks
Acelpryn (chlorantraniliprole) Enfold (emmamectin benzoate)	77 to 100 %	2 to 3 weeks
Intrepid (methoxyfenozide) Scimitar (lambda-cyhalothrin)	94 to 100 %	3 to 4 weeks
Pure Spray Green (horticultural oil)	74 to 85 %	+ / -
Dimilin (difluobenzuron)	Not effective	

Treatments at maximum labeled rate. Dyne-Amic surfactant added. Results based on survival to adulthood (2 repeated experiments). **% Control:** treatments applied after egg laying. **Residual**: treatments applied before egg laying.

"Today light brown apple moth is successfully managedthrough a combination of biological control and threshold-based applications of selective insecticides"

from New Zealand lessons...... California Agriculture Vol 64 No.1

New Zealand lessons may aid efforts to control light brown apple moth in California

by Lucia G. Varela, James T.S. Walker, Peter L. Lo and David J. Rogers

REVIEW ARTICLE

New Zealand's major fruit industries are dependent upon producing highquality crops for export with a very low incidence of pest damage. Light brown apple moth was an economically important pest within the fruit sector in the 1960s through the 1980s, and it developed resistance to broad-spectrum insecticides. The increase in its pest status focused research on biological control, and existing native natural enemies were augmented with new introductions from Australia in the late 1960s. By the early 1990s, this effort resulted in substantially reduced leafroller populations and fruit damage. The implementation of integrated pest management (IPM) programs in the New Zealand fruit sector in the midto late 1990s practically eliminated the use of broad-spectrum organophosphate insecticides, further enhancing natural control. Today light brown apple moth is successfully managed in IPM and organic programs through a combination of biological control and threshold-based applications of selective insecticides.

The recent discovery of light brown apple moth, a leafroller, in California may affect the management of fruit crops, and because it is a quarantine pest in some markets, the discovery has already had implications for domestic and export trade in produce and nursery stock.

In New Zealand, light brown apple moth, Epiphyas postvittana (Walker) (Lepidoptera: Tortricidae), was first reported in 1891 (Hudson 1928). It became a major pest, primarily of pome

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In New Zealand, light brown apple moth is controlled by parasitoids and selective insecticides.

fruit (apples and pears) and berry fruit, and a minor pest of grapes, citrus, stone fruit and kiwi fruit. The number of acres planted to pome and berry fruit — the crops most affected by light brown apple moth — is similar in New Zealand and Californi (table 1).

New Zealand went through a phase from the 1960s to 1980s when light brown apple moth caused major fruit damage; in apple crops this averaged from 8% to 26% and as high as 48% (Collyer and van Geldermalsen 1975). Control programs were based on frequent applications of broad-spectrum insecticides. This led to the development of resistance to organochlorines in the early 1960s (Collyer and van Geldermalsen 1975) and organophosphates by the early 1980s (Suckling et al. 1984; Suckling and Khoo 1990).

Over the last two decades, the post status of light brown apple moth in New Zealand apples has shifted significantly. Damage has decreased to typically less than 2% in unsprayed trees (fig. 1). The decline in fruit damage is associated with lower leafroller density, which in turn is attributed to two key factors: (1) the introduction in the 1960s and subsequent spread of parasitoids attacking pupal and late larval stages for light brown apple moth and (2) the change in fruit production programs from frequent applications of broadspectrum insecticides to less-intensive spraying with selective products.

By using a combination of natural control and selective insecticides, New Zealand growers are able to control light brown apple moth and meet the export standards of more than 60 countries that import a variety of fruit crops. In the 1980s, organophosphate insecticides were sprayed in six to nine applications each season on pome fruit crops for a variety of pests. Over the last decade, use of organophosphate insecticides has declined by 97%, while the frequency of insecticide applications has declined by approximately 50% (Manktelow et al. 2005). The insecticides now used are selective. The incidence of light brown apple moth fruit damage has declined, as has the larval incidence in crops.

In recent U.S. Department of Agriculture preclearance inspections of New Zealand apples grown using the Integrated Fruit Production program, the rejection rate of export consignments for the presence of light brown apple moth was typically less than 1%. A consignment is rejected if one or more larva is detected in 20,000 individually inspected fruit.

Biology and damage

The light brown apple moth's biology was previously described in *California Agriculture* (Varela et al. 2008). In New Zealand, this insect reportedly feeds on 265 different host plant species

TABLE 1. Planted area of light brown apple moth fruit-crop hosts			
Сгор	New Zealand*	Californiat	
	· · · · · · · acres	(hectares) · · · · · ·	
ome fruit	23,539 (9,526)	36,500 (14,771)	
lerry fruit	5,913 (2,393)	7,400 (2,994)	
trawberries	420 (170)	7,400 (2,994)	
rapes	72,518 (29,347)	35,500 (14,366)	
itrus	4,532 (1,834)	789,000 (319,297)	
one fruit		251,500 (101,778)	
wi fruit	5,669 (2,294)	243,800 (98,662)	
	30,112 (12,186) search Fresh Facts 200	4 000 (1 510)	